FVSP Faculty Application Checklist-DUE 1/19/2024 5PM EST

X Completed cover page with prior mentorship history

X Training/Registration requirements needed

X Abstract of proposed work

X NIH-format biosketch

Submission Instructions

Convert the application to ***one .pdf document.*** Name the file using your last name, followed by an underscore, and your first initial. For example: Martyniuk_C.pdf

Submit the following pages, via email attachment, to Dr. Chris Martyniuk (cmartyn@ufl.edu). The subject line should read "FVSP Faculty Application".

The FVSP Research Program runs 5/27/2024 to 8/07/2024 with final research presentations prior to the national symposium.

2024 Linda F. Hayward Florida Veterinary Scholars Program
Faculty Application

Name	Francesca Griffin
Email address	fcgriffin@ufl.edu
Proposed project title	Analysis and Culture Results of Spontaneously Voided Urine Samples in Cats: An Observational Study
Will you provide matching student stipend funding ($3250)?	I am hoping to provide matching funding, though I don't have a source yet
Source of project/research funding	

Prior student research mentees (last 5 years, if applicable):

CLASS	STUDENT	PROJECT TITLE	STATUS
2021	Alyssa Blew (FVSP 2018)	A fear free approach to investigating heart rate fluctuations during veterinary examinations in varying locations as an indicator of stress in dogs	Published in JSAP
2021	Alexis Deriberprey (FVSP 2018)	Effects of physical exam location on heart rate and outward manifestation of signs of fear, anxiety, and stress in cats	Published in JFMS
2024	Monica Suero (FVSP 2021)	Analysis and culture results of free catch urine sample in dogs	Submitted to JSAP – under review

If project qualifies for Morris Animal Foundation Student Scholarship Funding and you have identified a specific interested student, please provide their name and email address

LAST NAME	FIRST NAME	EMAIL ADDRESS
N/A		

I agree to obtaining all necessary approvals (e.g. IACUC/IRB/EH&S/VHRRC – see below for specifics) to conduct the project with the student PRIOR to the commencement of the summer program, as well as submitting documentation of these approvals to the FVSP board by 5/11/2024

YES

I agree to assisting my student prepare for the summer program during the Spring semester, which will include preparation of a study outline, and training in relevant laboratory techniques

YES

I agree to plan for commencing the experiment/data collection by the beginning of the summer program (5/22/24)

YES

I agree to be available to the student throughout the summer to assist with the experiment/data collection, preparation of the manuscript and poster.

YES

	Needed (Yes/No)	Approval by 5/11/24 (Yes/No)?
IACUC Approval and Training	No – see attached letter	No
IRB Registration and Training	No	No
Biological Agent Registration	No	No
Biopath Registration	No	No
Veterinary Hospital Research	Done – VHRRC # 2023-16	Yes
FERPA Training	No	No
Biohazardous Waste Training	No	No
Laboratory Safety Training	No	No

Abstract of proposed student project (1 page limit. This should mirror the aims page of a grant and CLEARLY indicate the student's role.)

Urine analysis is a very common and important test performed on a daily basis in a small animal general practice setting. Among other uses, urine analysis is used to evaluate kidney function, monitor diabetic patients, assess protein loss, and is the primary diagnostic tool used to evaluate for bacterial urinary tract infection in cats presenting with urinary signs. Because urine analysis is an important tool in diagnosing and treating many disease processes, it is considered part of a minimum data base of lab work recommended for both sick patients and patients presenting for annual wellness evaluation.

Urine can be collected from feline patients in 3 ways: cystocentesis, urethral catheterization, and spontaneous voiding. Spontaneous voiding, while least invasive, poses a logistical challenge in feline patients as the majority are not trained to void upon command. Additionally, most felines will not urinate while in a veterinary hospital setting unless they are confined for an extended period of time. Hence, cleaning the external genitalia and collecting a mid-stream free catch sample is typically not feasible in feline patients. An alternative method for urine collection in cats involves substituting standard absorbent litter substrates with a non-absorbable, pelleted substrate so that urine can be collected from the litter box after spontaneous voiding. Owners can be sent home with the pelleted substrate and directions for appropriate sample collection and storage, and then returning the sample to the veterinary hospital for analysis. Proper protocol entails cleaning the litter box with dish soap and hot water, drying it thoroughly, placing the substrate in the box, and frequently checking the box for urine. Once urine is produced, they can use a sterile syringe to collect the urine and seal with a sterile syringe cap. If the owners cannot return the urine to the hospital immediately, then they will need to refrigerate the sample. The sample should be fully processed within 24 hours of collection to minimize artifact formation (casts, crystals, pH changes, degradation of cells) or ex vivo bacterial contamination. If it can be determined that bacterial contamination occurs infrequently with appropriate spontaneous voiding collection methods, this low-risk method may be considered a more acceptable option for urine collection in cats than more invasive procedures.
This will be an observational study, as feline participants will not be entering the UF Small Animal Hospital. Instead, the owners will pick up a home collection kit and bring the urine samples to the hospital once obtained.

A urinalysis and urine culture will be performed on all samples. The primary outcome is presence/absence of contamination defined as $>10^4$ CFU/mL mixed flora, as identified by the veterinary clinical laboratory. Spontaneously voided samples that are positive for hematuria, bacteriuria, or pyuria may require the patient to have a sterile sample collected via cystocentesis and submitted to the diagnostic lab if they have clinical signs of urinary tract disease.

The goal is to recruit 100 feline participants. Patient exclusionary criteria are those with systemic illness that predispose them to secondary urinary tract infections: diabetes mellitus, hyperthyroidism, GI disease, and chronic kidney disease.

The students assigned to our project will be responsible for identification and recruitment of participants, obtaining study consent, and distribution of sample collection kits. Once returned to the hospital, the students will evaluate the urine samples (wet and dry mount urinalyses) and plate for culture. The objective is to analyze the results and submit a manuscript to a peer-reviewed journal.

OMB No. 0925-0001 and 0925-0002 (Rev. 11/16 Approved Through 10/31/2018)

BIOGRAPHICAL SKETCH

Provide the following information for the Senior/key personnel and other significant contributors.
Follow this format for each person. **DO NOT EXCEED FIVE PAGES.**

NAME: Francesca Corbly Griffin

eRA COMMONS USER NAME (credential, e.g., agency login): N/A

POSITION TITLE: Clinical Assistant Professor

EDUCATION/TRAINING *(Begin with baccalaureate or other initial professional education, such as nursing, include postdoctoral training and residency training if applicable. Add/delete rows as necessary.)*

INSTITUTION AND LOCATION	DEGREE *(if applicable)*	Completion Date MM/YYYY	FIELD OF STUDY
Rollins College	BA	05/1993	Biology
University of South Florida College of Medicine – Tampa, FL	MS	12/1997	Biomedical Sciences
University of Florida College of Veterinary Medicine – Gainesville, FL	DVM	05/2003	Small Animal Focus

A. Personal Statement

I have been a veterinarian for 20 years, and during that time, I have worked in both a private practice setting and in academia. During that time, I have cared for thousands of animals, and have observed that patient stress / anxiety is a significant concern for both owners and veterinary personnel. Patient stress, and its manifestations, can be a significant factor in poor owner compliance, with regards to routine exams, diagnostics and therapies. Since two of my central goals in practicing veterinary medicine are to improve my patient's quality of life and strengthen the bond between owners and their pets, I have always strived to do what I can to minimize the angst animal's experience while in my care. That is why I became interested in the Fear Free Pets Initiative, and became certified in September 2017. The initiative covers techniques for minimizing stress in animals, and client communication is a very important component. Both my training through the Bayer Communications Project and experience in communication rounds with students, residents, and house officers have helped to strengthen these skills. Since completing the Fear Free training, I have implemented a number of techniques into the daily care of my patients, and both owners and animals appear to appreciate the extra effort, and

appear less stressed. Prior research projects entailed evaluating exam location in the hospital and its contribution of stress in felines and canines, as evidenced by changes in heart rate. I have also participated in an FVSP study that evaluated the degree of bacterial contamination in free catch urine samples in canines. I'm now turning my attention to evaluating the diagnostic utility of free catch urine samples, collected at home, in feline patients.

B. Positions and Honors

Positions & Employment:

- Associate Veterinarian Millhopper Veterinary Medical Center June 2003 – September 2013
- Relief Small Animal Veterinarian September 2013 – November 2014
- Clinical Assistant Professor UF-CVM November 2014 – present

Veterinary Affiliations & Activities:

- UF-CVM Admissions Committee Member 2023 – 2026 (3-year commitment)
- UF-CVM Search Committee Member for Dermatology Faculty Candidate 2022 – 2023
- UF-CVM Representative on Southeast Veterinary Education Consortium Steering Committee 2022 – present
- UF-CVM Curriculum Review Committee Member 2018 – 2019
- UF-CVM Student-Faculty Communications Committee Chair 2017 – 2019
- UF-CVM Student-Faculty Communications Committee Member 2015 – 2021
- UF-CVM House Officer Program Review Committee Member 2016 – 2018
- UF-CVM Search Committee Member for Primary Care & Dentistry Clinical Lecturer Candidate 2017
- Lecturer UF Premed AMSA's Annual Health Career Convention 2/2016, 2/2018, 2/2019, 2/2021, 4/2022 (invited but meeting canceled by coordinators at last minute)
- Lecturer UF-CVM Annual Open House 4/2016
- American Veterinary Medical Association Member since 2003
- Alachua Veterinary Medical Association Member since 2003
- Alachua VMA President 2015-16 and 2020-21
- Alachua VMA Board Member 2014 – 2017, 2019 – 2021
- American Heartworm Society Member since 2019
- Association of American Veterinary Medical Colleges Member since 2014
- Primary Care Veterinary Educators Member since 2014
- Florida Veterinary Medical Association Member 2003 – 2007, 2017 – present
- FVMA Conference Attendee 2017 – 2018
- NAVC Attendee 2016, 2015, 2014, 2012, 2007, 2006, 2005
- UF-CVM Visiting Practitioner Program in SA Medicine 2009
- Attendee 19th Annual Veterinary Dental Forum 2005
- Attendee UF-CVM Referring Veterinarian Appreciation Day 2007, 2008, 2013
- USDA Accredited Veterinarian since 2003
- UF-CVM Class of 2003 President 2002 – 2003
- UF-CVM Class of 2003 Secretary 2001 – 2002 Veterinary Medicine College Council Student Representative 2000 – 2001

- UF-CVM Curriculum Committee Class Representative 1999 – 2003
- Student Chapter of American Veterinary Medical Association 1999 – 2003

Honors:

- FVMA Gold Star Award Recipient 2020
- Delivered UF-CVM Class of 2003 student commencement address
- Toby Fastiggi Award Recipient 2003
- Allan H. Hart / IDEXX Scholarship Recipient 2003
- Merck Veterinary Manual Award Recipient 2003
- Member of *Phi Zeta* Veterinary Honor Society since May 2002
- University of Florida Presidential Recognition Award 2001
- American Society of Andrology Thomas K. Chang Award Recipient 1997
- University of South Florida University Graduate Fellowship Recipient 1996
- Member of *Phi Kappa Phi* Honor Society since 1995
- Dean's List - Rollins College - Fall 1990 and Spring 1993

C. Contributions to Science

While I was a graduate student at USF College of Medicine, my area of research was studying the role of Fas / FasL in the testes of rats. I worked on developing a rat model, where blood flow to the testes was transiently occluded with a vascular clip. Immunohistochemistry markers were used to identify the anatomical location, and potential up regulation, of Fas / FasLin the testes of normal and abnormal testes (i.e. blood flow obstruction).

"Immunoexpression of Fas (CD95) Ligand (FasL) in the Rat Testis and Epididymis", F.C. Griffin & D.F. Cameron. Presented at the 22nd Annual Meeting of the American Society of Andrology, February 22-25, 1997, Baltimore, MD

"Localization and Expression of Fas (CD95) Ligand (FasL) and Fas Receptor (Fas) in the Rat Epididymis", F.C. Griffin, D.A. Lowe, T. Lund & D.F. Cameron. Presented at Experimental Biology Meeting'97, April 6-9, 1997, New Orleans, LA.

"Immunoexpression of Fas (CD95) Ligand (FasL) in the Leydig Cells of Normal and Hypophysectomized Rats", F.C. Griffin, J. Hushen, P. Barbour & D.F. Cameron. Presented at the 23rd Annual Meeting of the American Society of Andrology, March 26-29, 1998, Long Beach, CA

"Ultrastructure of Sertoli - Germ Cell Interactions in the Normal and Pathologic Testis", D.F. Cameron, F.C. Griffin. Chapter 18 of Male Reproduction: A Multidisciplinary Overview, Churchill Communications Europe Espana, 1998.

While employed at the UF College of Veterinary Medicine, my area of focus has been teaching veterinary students, as they rotate through the Primary Care & Dentistry Service. During that time, I have participated in clinical research, some of which has been published.

"Diagnostic Imaging in Veterinary Dental Practice", J. Wuerz, J. Rooks, & **F. Griffin.** Published in *Journal of American Veterinary Medical Association.* Published on-line June 22, 2022. Published in Volume 260, issue 14, pp. 1789-1792, November 2022. https://doi.org/10.2460/javma.22.04.0160

"Evaluation of Clinical Exam Location on Stress in Cats: A Randomized Crossover Trial", **F. Griffin**, W. Mandese, P. Reynolds, A. Deriberprey, & A. Blew. Published in *Journal of Feline Medicine & Surgery.* Published on-line October 15, 2020. Published in Volume 23, issue 4, pp. 364-369, April 2021. *JFMS* Editor's top-pick article for April 2021. https://doi.org/10.1177/1098612X20959046

"Stress in Client-Owned Dogs Related to Clinical Exam Location: A Randomised Crossover Trial", W. Mandese, **F. Griffin**, P. Reynolds, A. Blew, & A. Deriberprey. Published in *Journal of Small Animal Practice.* Published on-line October 26, 2020. Published in Volume 62, issue 2, pp. 82-88, February 2021. https://doi.org/10.1111/jsap.13248

"What is Your Diagnosis? Peripheral Blood Smear and Splenic Fine Needle Aspirate from a Cat", F. Conrado, J. Stern & **F. Griffin**. Published *Veterinary Clinical Pathology*. Published on-line February 19, 2020. Published in Volume 49, issue 2, pp. 367-370, June 2020. https://doi.org/10.1111/vcp.12817

"Overview of a Practice Based Rotating Small Animal Clerkship", W Mandese, F. Griffin, LS Behar-Hornstein. Presented at Veterinary Educators Conference, June 2016.

"The Impact of a Wellness Centered Approach in Practice Growth within a University Primary Care Setting", A.E.S. Stone, C.M.P. Hermansen, J.A. Wuerz, W. Mandese & F. Griffin. Presented at Primary Care Veterinary Educators World Symposium, October, 2015.

D. Additional Information: Research Support and/or Scholastic Performance:
Fear Free grant for summer 2018 FVSP feline project